Highest Recorded N−O Stretching Frequency for 6‑Coordinate {Fe- $NO⁷$ Complexes: An Iron Nitrosyl Model for His₃ Active Sites

Jia Li,† Atanu Banerjee,† Piotr L. Pawlak,† William W. Brennessel,‡ and Ferman A. Chavez*,†

† Department of Chemistry, Oakland University, Rochester, Michigan 48309-4477, United States

‡ Department of Chemistry, University of Rochester, Rochester, New York 14627-0216, United States

S Supporting Information

[AB](#page-2-0)STRACT: [We](#page-2-0) [report](#page-2-0) [t](#page-2-0)he synthesis, structure, and reactivity of $[Fe(T1Et4iPrIP)(OTf)_2]$ $[1; T1Et4iPrIP =$ tris(1-ethyl-4-isopropylimidazolyl)phosphine]. Compound 1 reacts reversibly with nitric oxide to afford [Fe- $(T1Et4iPrIP)(NO)(THF)(OTf)(OTf)$ (2), which is the first example of a 6-coordinate $\{FeNO\}^7$ $S = \frac{3}{2}$ complex containing a linear Fe−N−O group. 2 exhibits the highest $\nu(NO)$ for compounds in this class. Density functional theory studies reveal an enhanced degree of β -electron transfer from $\pi^*(NO)$ to the Fe d orbitals accounting for the large stretching frequency.

C lass III nonheme dioxygenases possess iron(II) coordina-
tion sites bearing three histidine residues, which contrasts
with Class I and II sites containing 2 His Lonbourlate matel with Class I and II sites containing 2His-1-carboxylate metal binding sites. The lack of a carboxylate group in the first coordination sphere renders the iron site more Lewis acidic by altering the mechanism for dioxygenase reactivity. Enzymes containing $His₃$ metal binding sites include gentisate 1,2dioxygenase, salicylate 1,2-dioxygenase, 3-hydroxyanthranilate 3,4-dioxygenase, 1-hydroxy-2-naphthoate dioxygenase, and cysteine dioxygenase.¹ These enzymes belong to the cupin superfamily characterized by two highly conservative sequence motifs of $G(X)_{5}HXH(X)_{3,4}E(X)_{6}G$ $G(X)_{5}HXH(X)_{3,4}E(X)_{6}G$ $G(X)_{5}HXH(X)_{3,4}E(X)_{6}G$ and $G(X)_{5}PXG(X)_{2}H (X)_{3}N^{2}$ Nitric oxide has been used to characterize ferrous active sites because they are electron paramagnetic resonance (EPR) sile[n](#page-2-0)t and do not exhibit low-energy absorption bands.³ Although many nonheme enzymes form stable Fe-NO complexes, a number of them [a](#page-2-0)re known to react reversibly $3a$ with the formation of paramagnetic nitrosyl iron centers, which according to the Enemark and Feltham notation are of the ${FeNO}^7$ ${FeNO}^7$ ${FeNO}^7$ type.⁵ These nonheme iron enzyme nitrosyl derivatives invariably possess a[n](#page-2-0) EPR-active $S = \frac{3}{2}$ ground state characterized by an axial EPR spectrum with g_{x} , g_{y} , and g_{z} values of ~4, ~4, and ~2, respectively. Researchers have used Raman, magnetic circular dichroism, EPR, Mössbauer spectroscopies, SCF-Xa-SW, and generalized gradient approximation (GGA) density functional theory (DFT) calculations and have determined that these {FeNO}⁷ (S = $\frac{3}{2}$) species are the result of high-spin ferric (S = $\frac{5}{2}$) antiferromagnetically coupled to NO⁻ (S - 1)^{3b,6} The $^{5}/_2$) antiferromagnetically coupled to NO[−] (S = 1).^{3b,6} The acquisition of synthetic complexes capable of reversibly binding NO is an important aspect for accurately modeli[ng](#page-2-0) these enzymes. In our efforts to synthesize models that reproduce the reversibility of iron nitrosyl formation, we have employed the ligand tris(1-ethyl-4-isopropylimidazolyl)phosphine (T1Et4iPrIP).⁷ The reaction of T1Et4iPrIP with $Fe(OTf)_{2}$: 2MeCN affords $[Fe(T1Et4iPrIP)(OTf)₂]$ (1; Figure 1). In the structure

Figure 1. (a) T1Et4iPrIP and (b) a thermal ellipsoid plot (50% probability) for 1. H atoms have been omitted for clarity.

of 1, iron(II) is bonded to three imidazole N atoms in a facial manner along with two O atoms from the triflates to afford highly distorted trigonal-bipyramidal geometry (τ = 0.33)⁸ with one O atom and one N atom occupying the apical positions.

The reaction of a colorless solution of 1 in tet[ra](#page-2-0)hydrofuran (THF) with nitric oxide at 25 °C yields a yellow-brown solution (Figure S1 in the Supporting Information, SI). The three-band pattern in the UV−vis spectrum, with relatively intense absorption featur[es in the near-UV regi](#page-2-0)on, followed by an absorption band with intermediate intensity in the 400−500 nm region and a weaker absorption band between the 600 and 700 nm region is characteristic of iron nitrosyl compounds.⁹ The band centered at ∼455 nm is assigned to a Fe−NO metal-toligand charge-transfer band. When a vacuum is applied [to](#page-2-0) the reaction mixture or argon is bubbled into the solution, the band near 455 nm diminishes, giving a spectrum similar to that of the starting material. This process is reversible over several cycles.

The crystal structure for the nitrosyl adduct reveals a bound NO molecule along with a coordinated THF and triflate group. The asymmetric unit cell contains two iron cations and two triflate groups. One of the cations (Figure 2) shows a slightly bent Fe−N−O unit with an angle of 168.6(5)° and Fe−NO and N−O distances of 1.765(5) and 1.146(6) Å[, r](#page-1-0)espectively, while the other exhibits a more linear Fe−N−O unit with an angle of 174.4(4)°, a Fe−NO distance of 1.763(5) Å, and a N−O

Received: March 17, 2014 Published: May 19, 2014

Figure 2. Thermal ellipsoid plot (50% probability) for [Fe- $(T1Et4iPrIP)(THF)(OTf)(NO)]$ ⁺ (cation of 2). H atoms have been omitted for clarity.

distance of 1.153(6) Å. The observed Fe−N−O angles can mostly be ascribed to electronic and not steric effects because the nearest interaction between the O atom of NO and the ligand is 3.174 Å (O1−C12). Having Fe−N−O angles greater than 165° constitutes 2 as the first example of a linear FeNO complex in this class.¹⁰

The EPR spectrum for 2 in toluene/THF glass at 77 K shows S $=$ $\frac{3}{2}$ [sig](#page-2-0)nals with effective g values of 3.90 and 2.01 (Figure S2 in the SI). These data are in accordance with the well-established electronic structure of nonheme ferrous nitrosyls, which show Fe^{III}–NO⁻ ground states where the high-spin Fe^{III} and NO⁻ (S = 1) are antiferromagnetically coupled. $3⁵$ The high concentration (20 mM) needed to achieve a good signal suggests a low concentration of this species, howev[er.](#page-2-0)

Although 2 is relatively stable in a THF/pentane solution at 25 [°]C under an atmosphere of NO, Park et al.¹¹ observed that [Fe(PhTIP)(acac)(NO)] [PhTIP = tris(2-phenylimidazol-4 yl)phosphine] was only moderately stable at −[40](#page-2-0) °C in MeCN. The difference in the nitrosyl-adduct stability could be due to either the higher steric shielding and/or greater electronwithdrawing nature of the 4-phenyl groups on PhTIP or the lack of a 1-alkyl group on the imidazole ring (compared to T1Et4iPrIP). Like 2, $[Fe(6TLA)(BF)](ClO₄)^{9a} {6TLA =$ tris $[(6-methyl-2-pyridyl)$ methyl]amine; BF = benzoylformate} reversibly binds NO to afford the nitrosyl adduc[t \[](#page-2-0)Fe(6TLA)- $(BF)(NO)[ClO₄)$.

Vibrational spectroscopy was used to further characterize the nature of the Fe−NO unit in 2 (Figure S3 in the SI). Importantly, the IR spectra of 2 (KBr) exhibits a $\nu(NO)$ stretching frequency, which is the highest (1831 cm⁻¹) observed [for](#page-2-0) 6-coordinate iron(II) nitrosyl complexes with ${FeNO}^7$ (Figure 3 and Table S1 in the SI), occurring only 44 cm[−]¹ below the stretching vibration of free NO (1875 cm^{-1})!¹² When the spectrum is taken in a THF s[olu](#page-2-0)tion, we observe a value of 1820 $\rm cm^{-1}$, suggesting that crystal packing may have so[me](#page-2-0) influence on the solid-state value.

There is a positive correlation between the Fe−N−O bond angle for 6-coordinate ${FeNO}^7$ complexes and $\nu(NO)$ (Figure 3).3b,9a,13−²⁰ The average Fe−N−O angle (171.5°) for 2 is used in this plot. Such a correlation was previously observed for 5 coordinate ${FeNO}$ ⁷ complexes.¹⁵ The correlation is particularly strong when compounds exhibiting spin-crossover^{15,22} and close contacts between the coordinat[ed](#page-2-0) NO and other ligands $6a,21$ are excluded.

Figure 3. Plot of the Fe–N–O angle versus $\nu(NO)$ for 6-coordinate ${[FeNO]}^7$ compounds (linear fit, $R = 0.95$).^{3b,9a,13–20}

In a study of 5-coordinate complexe[s,](#page-2-0) $2³$ $2³$ $2³$ [th](#page-2-0)e [re](#page-2-0)lationship seen in Figure 3 was reversed. One explanation for this could be that the observed Fe−N−O angle was not u[nde](#page-2-0)r electronic but steric control. The interaction between the NO group and ligand substituents could influence $\nu(NO)$, thus masking the correlation. It can be seen that the least influenced NO ligand in this study along with a recent example 24 correlates well with the previously mentioned study.¹⁵

DFT calculations have previously bee[n p](#page-2-0)erformed on FeNO compounds.^{9b,24} The PBE0 6-31[+G](#page-2-0)^{*}-optimized structure on the cation of 2 shows excellent agreement with experimental values. The Fe−N[O dis](#page-2-0)tances of 1.765 and 1.789 Å are only slightly smaller than the calculated value of 1.795 Å. Fe−N−O angles of 164.5 and 174.4°are very close to the calculated value of 173.24°. The N−O bond distances of 1.146 and 1.153 Å (calcd: 1.157 Å) also compare well. Most importantly, the calculated N−O stretching frequency of 1839 cm⁻¹ (scale factor = 0.938) is very close to the experimental (KBr) value (1831 cm[−]¹). Thus, the DFT results essentially reproduce the experimental metric and vibrational values. Analysis of the molecular orbital (MO) diagram demonstrates an electronic-structural description that is consistent with the high-spin Fe^{III}-NO⁻ bonding scheme.^{3b} Correspondingly, in the α -spin MO diagrams, all of the Fe d orbitals are (singly) occupied. In the applied coordinate syste[m](#page-2-0) (Fe–NO vector corresponding to the z axis), the d_{xz} and d_{yz} orbitals do not participate in back-bonding with the two unoccupied $\alpha-\pi^*$ orbitals of NO.

In the $β$ -spin MO diagram, all of the Fe d orbitals are empty, whereas the $\beta - \pi^*$ orbitals of NO are occupied. These occupied orbitals are situated to donate into the empty β -spin d_{xz} and d_{vz} orbitals of Fe. The strength of this interaction can be estimated from the corresponding antibonding combinations β 193 (42%) d_{yz} 58% π^*) and β194 (38% d_{xz} , 62% π^*) for the cation of 2, which show significant Fe d orbital and NO π^* character (see Figure 4). As a result, the π donation from NO⁻ is significant. This result was also observed in $[Fe(BMPA-Pr)Cl(NO)]^{13}$

The [c](#page-2-0)alculated spin-density values for Fe and NO are +3.93 and −1.15, respectively. These values reflect the strong do[nat](#page-2-0)ion of negative (β) spin density from NO to Fe, rendering NO to appear closer to NO^o and Fe closer to Fe²⁺. Because the donation originates from N−O π ^{*} orbitals, a strengthening of this interaction should result in a strengthening of the N−O bond and a corresponding increase in the N−O stretch. The description of NO as closer to that of NO• could account for the greater reactivity compared to that of [Fe(BMPA-Pr)Cl- (NO)]. Additionally, Lehnert and co-workers observed that

Figure 4. β -spin isosurface plots for Fe d orbitals and π^* orbitals of NO for the cation of 2: (a) β 193 (Fe 42%, NO 58%); (b) β 194 (Fe 42%, NO 58%).

decreasing the negative charge on the iron center results in increasing the iron's ability to accept electron donation from bound NO[−]. ¹³ Unlike [Fe(BMPA-Pr)Cl(NO)], we do not observe back-bonding between Fe $\alpha - d_z^2$ and $\alpha - \pi^*$ of NO⁻ because of the near-linearity of the Fe−N−O bond in 2. The lack of α -spin donation from Fe to NO could also account for the long Fe−NO bond distance.

In summary, we have synthesized an accurate model for $His₃$ enzymes. The model reversibly binds nitric oxide to generate a 6 coordinate ${FeNO}^7$ iron nitrosyl species with a nearly linear Fe−N−O bond angle. The $\nu(NO)$ stretching vibration is the highest recorded to date, and a strong correlation between the Fe−N−O bond angle and $\nu(NO)$ is observed when these parameters are plotted for other known 6-coordinate ${FeNO}$ complexes. Computational studies indicate that the bonding in the nitrosyl group can be described as a high-spin $\text{Fe}^{\text{III}} \text{ (}S = {}^{5}/_{2}\text{)}$ antiferromagnetically coupled to NO^{$-$} (S = 1), where the Fe− NO bond consists mostly of β electrons being donated back to the iron center. This accounts for the high N−O vibrational frequency.

■ ASSOCIATED CONTENT

6 Supporting Information

Experimental materials, methods, and figures along with crystallographic information (CIF). This material is available free of charge via the Internet at http://pubs.acs.org.

■ AUTHOR INFORMATION

Corresponding Author

*E-mail: chavez@oakland.edu.

Notes

The aut[hors declare no compe](mailto:chavez@oakland.edu)ting financial interest.

■ ACKNOWLEDGMENTS

We thank Prof. M. D. Sevilla for assistance with EPR. We thank Dr. V. G. Young, Jr. for X-ray structure determination of 1. We thank Prof. M. M. Szcześniak for assistance with the DFT calculations. National Science Foundation (NSF) Grant CHE-0748607 is gratefully acknowledged. An NSF award (Grant CHE-0821487) is also acknowledged.

■ REFERENCES

(1) (a) Buongiorno, D.; Straganz, G. D. Coord. Chem. Rev. 2013, 257, 541. (b) Chavez, F. A.; Banerjee, A.; Sljivic, B. Modeling the Metal Binding Site in Cupin Proteins. In On Biomimetics; Pramatarova, L. D., Ed.; InTech: Rijeka, Croatia, 2011.

(2) Dunwell, J. M.; Culham, A.; Carter, C. E.; Sosa-Aguirre, C. R.; Goodenough, P. W. Trends Biochem. Sci. 2001, 26, 740.

(3) (a) Orville, A. M.; Chen, V. J.; Krauciunas, A.; Harpel, M. R.; Fox, B. G.; Münck, E.; Lipscomb, J. D. Biochemistry 1992, 31, 4602. (b) Brown, C. A.; Pavlosky, M. A.; Westre, T. E.; Zhang, Y.; Hedman, B.; Hodgson, K. O.; Solomon, E. I. J. Am. Chem. Soc. 1995, 117, 715.

(4) (a) Arciero, D. M.; Lipscomb, J. D.; Huynh, B. H.; Kent, T. A.; Munck, E. J. Biol. Chem. 1983, 258, 4981. (b) Nocek, J. M.; Kurtz, D. M.; Sage, J. T.; Xia, Y. M.; Debrunner, P.; Shiemke, A. K.; Sandersloehr, J.; Loehr, T. M. Biochemistry 1988, 27, 1014. (c) Rodriguez, J. H.; Xia, Y. M.; Debrunner, P. G. J. Am. Chem. Soc. 1999, 121, 7846. (d) Haskin, C. J.; Ravi, N.; Lynch, J. B.; Mü nck, E.; Que, L. Biochemistry 1995, 34, 11090. (e) Chen, V. J.; Orville, A. M.; Harpel, M. R.; Frolik, C. A.; Surerus, K. K.; Mü nck, E.; Lipscomb, J. D. J. Biol. Chem. 1989, 264, 21677.

(5) Enemark, J. H.; Feltham, R. D. Coord. Chem. Rev. 1974, 13, 339. (6) (a) Hauser, C.; Glaser, T.; Bill, E.; Weyhermuller, T.; Wieghardt, K. J. Am. Chem. Soc. 2000, 122, 4352. (b) Westre, T. E.; Dicicco, A.; Filipponi, A.; Natoli, C. R.; Hedman, B.; Solomon, E. I.; Hodgson, K. O. J. Am. Chem. Soc. 1994, 116, 6757.

(7) Lynch, W. E.; Kurtz, D. M., Jr.; Wang, S.; Scott, R. A. J. Am. Chem. Soc. 1994, 116, 11030.

(8) Addison, A. W.; Rao, T. N.; Reedijk, J.; van Rijn, J.; Verschoor, G. C. J. Chem. Soc., Dalton Trans. 1984, 1349.

(9) (a) Chiou, Y. M.; Que, L. Inorg. Chem. 1995, 34, 3270. (b) Berto, T. C.; Speelman, A. L.; Zheng, S.; Lehnert, N. Coord. Chem. Rev. 2013, 257, 244.

(10) McCleverty, J. A. Chem. Rev. 2004, 104, 403.

(11) Park, H.; Bittner, M. M.; Baus, J. S.; Lindeman, S. V.; Fiedler, A. T. Inorg. Chem. 2012, 51, 10279.

(12) Richter-Addo, G. B.; Legzdins, P. Metal Nitrosyls; Oxford University Press: New York, 1992.

(13) Berto, T. C.; Hoffman, M. B.; Murata, Y.; Landenberger, K. B.; Alp, E. E.; Zhao, J. Y.; Lehnert, N. J. Am. Chem. Soc. 2011, 133, 16714.

(14) Randall, C. R.; Zang, Y.; True, A. E.; Que, L.; Charnock, J. M.; Garner, C. D.; Fujishima, Y.; Schofield, C. J.; Baldwin, J. E. Biochemistry 1993, 32, 6664.

(15) Weber, B.; Gorls, H.; Rudolph, M.; Jager, E. G. Inorg. Chim. Acta 2002, 337, 247.

(16) Rose, M. J.; Patra, A. K.; Olmstead, M. M.; Mascharak, P. K. Inorg. Chim. Acta 2010, 363, 2715.

(17) Nebe, T.; Beitat, A.; Wurtele, C.; Ducker-Benfer, C.; van Eldik, R.; McKenzie, C. J.; Schindler, S. Dalton Trans. 2010, 39, 7768.

(18) McQuilken, A. C.; Ha, Y.; Sutherlin, K. D.; Siegler, M. A.; Hodgson, K. O.; Hedman, B.; Solomon, E. I.; Jameson, G. N.; Goldberg, D. P. J. Am. Chem. Soc. 2013, 135, 14024.

(19) Lopez, J. P.; Heinemann, F. W.; Prakash, R.; Hess, B. A.; Horner, O.; Jeandey, C.; Oddou, J. L.; Latour, J. M.; Grohmann, A. Chem.-Eur. J. 2002, 8, 5709.

(20) Patra, A. K.; Rowland, J. M.; Marlin, D. S.; Bill, E.; Olmstead, M. M.; Mascharak, P. K. Inorg. Chem. 2003, 42, 6812.

(21) Pohl, K.; Wieghardt, K.; Nuber, B.; Weiss, J. J. Chem. Soc., Dalton Trans. 1987, 187.

(22) Li, M.; Bonnet, D.; Bill, E.; Neese, F.; Weyhermüller, T.; Blum, N.; Sellmann, D.; Wieghardt, K. Inorg. Chem. 2002, 41, 3444.

(23) Ray, M.; Golombek, A. P.; Hendrich, M. P.; Yap, G. P. A.; Liable-Sands, L. M.; Rheingold, A. L.; Borovik, A. S. Inorg. Chem. 1999, 38, 3110.

(24) Speelman, A. L.; Lehnert, N. Angew. Chem., Int. Ed. 2013, 52, 12283.